

## Резюме: Мартынов Владимир Иванович

### Адрес

Федеральное государственное бюджетное учреждение науки Институт биоорганической химии им. академиков М.М. Шемякина и Ю.А. Овчинникова Российской академии наук, Москва, Россия

### Контакты

<https://www.ibch.ru/ru/users/38>

### Образование

1983–2013	Россия, Москва	Институт биоорганической химии им. академиков М.М. Шемякина и Ю.А. Овчинникова РАН (ИБХ РАН)	Присуждена учёная степень доктора химических наук, диссертация «Посттрансляционные модификации белков семейства GFP»
1975–1983	Россия, Москва	Институт биоорганической химии им. М.М. Шемякина АН СССР (ИБХ)	Присуждена учёная степень кандидата химических наук, диссертация «Исследование молекулярной организации зрительного родопсина »
1970–1975	Россия, Москва	Московский государственный университет им. М.В. Ломоносова (МГУ, химический факультет)	Диплом химика

### Работа в ИБХ

2018–наст.вр.	Главный научный сотрудник
---------------	---------------------------

### Научные интересы

Основные научные интересы В.И. Мартынова связаны с исследованиями структуры и функции светочувствительных белков, а также возможностью их применения в молекулярной, клеточной биологии и в тераностике различных заболеваний.

### Степени и звания

Доктор наук (Химические науки)
--------------------------------

### Гранты и проекты

2023–наст.вр.	<a href="#">Модульные пептидно-белковые конструкции для ранней диагностики и лечения онкологических заболеваний</a>
2020–2022	<a href="#">Гибридные конструкции для нацеленной доставки синтетических фотосенсибилизаторов к раковым клеткам</a>
2018–2020	<a href="#">Создание новых генетически кодируемых агентов для тераностики раковых заболеваний</a>

### Публикации

- Frolova AY, Pakhomov AA, Deyev SM, **Martynov VI** (2024). Influence of Peptide Linkers on the Functional Properties of Hybrid Structures with Selective pH-Dependent Binding to Cancer Cells. *Russ. J. Bioorganic Chem.* 50 (4), 1141–1149, [10.1134/S1068162024040010](#)
- Frolova AY, Kim EE, Kononevich YN, **Martynov VI**, Deyev SM, Pakhomov AA (2024). Octa-BODIPY dye for monitoring live cell parameters using fluorescence lifetime imaging microscopy. *Opt Spectrosc* 132 (4), 358–

363, [10.61011/EOS.2024.04.58874.26-24](https://doi.org/10.61011/EOS.2024.04.58874.26-24)

3. Frolova AY, Pakhomov AA, Kakuev DL, Sungurova AS, Dremina AA, Mamontova ED, Deyev SM, **Martynov VI** (2023). Hybrid protein-peptide system for the selective pH-dependent binding and photodynamic ablation of cancer cells. *J Photochem Photobiol B* 249, 112803, [10.1016/j.jphotobiol.2023.112803](https://doi.org/10.1016/j.jphotobiol.2023.112803)
4. Efimova AS, Ustimova MA, Maksimova MA, Frolova AY, **Martynov VI**, Deyev SM, Pakhomov AA, Fedorov YV, Fedorova OA (2023). Synthesis and optical characteristics of 4-styrylpyridinium dyes and their conjugates with antibody. *MENDELEEV COMMUN* 33 (3), 384–386, [10.1016/j.mencom.2023.04.027](https://doi.org/10.1016/j.mencom.2023.04.027)
5. Pakhomov AA, Efremova AV, Kononevich YN, Ionov DS, Maksimova MA, Volodin AD, Korlyukov AA, Dubinets NO, **Martynov VI**, Ivanov AA, Muzafarov AM (2023). NIR-I fluorescent probes based on distyryl-BODIPYs with two-photon excitation in NIR-II window. *ChemPhotoChem* 7 (5), e20220032, [10.1002/cptc.202200324](https://doi.org/10.1002/cptc.202200324)
6. Mamaeva AA, Frolova AY, Kakuev DL, **Martynov VI**, Deyev SM, Pakhomov AA (2023). Co-expression of different proteins in Escherichia coli using plasmids with identical origins of replication. *Biochem Biophys Res Commun* 641, 57–60, [10.1016/j.bbrc.2022.12.020](https://doi.org/10.1016/j.bbrc.2022.12.020)
7. Pakhomov AA, Efremova AV, Maksimova MA, Kononevich YN, Ionov DS, Dubinets NO, **Martynov VI**, Muzafarov AM, Alfimov MV (2023). Spectral Properties of Functional Distyryl Derivatives of BODIPY. *HIGH ENERG CHEM+* 57 (3), 192–199, [10.1134/S0018143923030104](https://doi.org/10.1134/S0018143923030104)
8. Frolova AY, Kutyakov SV, **Martynov VI**, Deyev SM, Pakhomov AA (2023). BODIPY Dye Derivative for Irreversible Fluorescent Labeling of Eukaryotic Cells and Their Simultaneous Cytometric Analysis. *Acta Naturae* 15 (4), 92–99, [10.32607/actanaturae.26879](https://doi.org/10.32607/actanaturae.26879)
9. Pakhomov AA, Belova AS, Khchoyan AG, Kononevich YN, Ionov DS, Maksimova MA, Frolova AY, Alfimov MV, **Martynov VI**, Muzafarov AM (2022). Ratiometric Singlet Oxygen Sensor Based on BODIPY-DPA Dyad. *Molecules* 27 (24), 9060, [10.3390/molecules27249060](https://doi.org/10.3390/molecules27249060)
10. Mamaeva AA, **Martynov VI**, Deyev SM, Pakhomov AA (2022). Comparison of Colorimetric and Fluorometric Chemosensors for Protein Concentration Determination and Approaches for Estimation of their Limits of Detection. *Chemosensors (Basel)* 10 (12), 542, [10.3390/chemosensors10120542](https://doi.org/10.3390/chemosensors10120542)
11. Frolova AY, Pakhomov AA, Kakuev DL, Sungurova AS, Deyev SM, **Martynov VI** (2022). Cancer cells targeting with genetically engineered constructs based on a pH-dependent membrane insertion peptide and fluorescent protein. *Biochem Biophys Res Commun* 612, 141–146, [10.1016/j.bbrc.2022.04.112](https://doi.org/10.1016/j.bbrc.2022.04.112)
12. Pakhomov AA, Kim EE, Kononevich YN, Ionov DS, Maksimova MA, Khalchenia VB, Maksimov EG, Anisimov AA, Shchegolikhina OI, **Martynov VI**, Muzafarov AM (2022). Modulation of the photophysical properties of multi-BODIPY-siloxane conjugates by varying the number of fluorophores. *Dyes Pigm* 203, , [10.1016/j.dyepig.2022.110371](https://doi.org/10.1016/j.dyepig.2022.110371)
13. **Martynov I**, Pakhomov A (2021). BODIPY derivatives as fluorescent reporters of molecular activities in living cells. *RUSS CHEM REV* 90 (10), 1213, [10.1070/RCR4985](https://doi.org/10.1070/RCR4985)
14. Goryashchenko AS, Pakhomov AA, Ryabova AV, Romanishkin ID, Maksimov EG, Orsa AN, Serova OV, Mozhaev AA, Maksimova MA, **Martynov VI**, Petrenko AG, Deyev IE (2021). FLIM-Based Intracellular and Extracellular pH Measurements Using Genetically Encoded pH Sensor. *Biosensors (Basel)* 11 (9), , [10.3390/bios11090340](https://doi.org/10.3390/bios11090340)
15. Orsa AN, Goryashchenko AS, Serova OV, Mozhaev AA, **Martynov VI**, Pakhomov AA, Svirshchevskaya EV, Petrenko AG, Deyev IE (2021). Generation and Characteristics of Genetically Encoded Fluorescent Sensors of Extracellular pH. *Russ. J. Bioorganic Chem.* 47 (5), 1060–1065, [10.1134/S1068162021040178](https://doi.org/10.1134/S1068162021040178)
16. Pakhomov AA, Pastukhova AA, Tishkin GV, **Martynov VI** (2021). Transformations of the Chromophore in the Course of Maturation of a Chromoprotein from Actinia equina. *Russ. J. Bioorganic Chem.* 47 (1), 230–235, [10.1134/S1068162021010167](https://doi.org/10.1134/S1068162021010167)
17. Frolova AY, Pakhomov AA, **Martynov VI** (2021). Physicochemical Properties of Photoconvertible Fluorescent Protein from Montastraea cavernosa. *Russ. J. Bioorganic Chem.* 47 (1), 244–251, [10.1134/S1068162021010052](https://doi.org/10.1134/S1068162021010052)
18. Pakhomov AA, Frolova AY, Tabakmakher VM, Chugunov AO, Efremov RG, **Martynov VI** (2020). Impact of external amino acids on fluorescent protein chromophore biosynthesis revealed by molecular dynamics and mutagenesis studies. *J Photochem Photobiol B* 206, 111853, [10.1016/j.jphotobiol.2020.111853](https://doi.org/10.1016/j.jphotobiol.2020.111853)
19. **Martynov VI**, Pakhomov AA, Deyev IE, Petrenko AG (2018). Genetically encoded fluorescent indicators for

- live cell pH imaging. *BIOCHIM BIOPHYS ACTA* 1862 (12), 2924–2939, [10.1016/j.bbagen.2018.09.013](https://doi.org/10.1016/j.bbagen.2018.09.013)
20. Pakhomov AA, **Martynov VI**, Orsa AN, Bondarenko AA, Chertkova RV, Lukyanov KA, Petrenko AG, Deyev IE (2017). Fluorescent protein Dendra2 as a ratiometric genetically encoded pH-sensor. *Biochem Biophys Res Commun* 493 (4), 1518–1521, [10.1016/j.bbrc.2017.09.170](https://doi.org/10.1016/j.bbrc.2017.09.170)
  21. Pakhomov AA, Mironiuk VB, Kononevich YN, Korlyukov AA, Volodin AD, Pryakhina TA, **Martynov VI**, Muzafarov AM (2017). Synthesis and crystal structure of a meso-decene-BODIPY dye as a functional bright fluorophore for silicone matrices. *MENDELEEV COMMUN* 27 (4), 363–365, [10.1016/j.mencom.2017.07.014](https://doi.org/10.1016/j.mencom.2017.07.014)
  22. Pakhomov AA, Chertkova RV, Deyev IE, Petrenko AG, **Martynov VI** (2017). Generation of photoactivatable fluorescent protein from photoconvertible ancestor. *Russ. J. Bioorganic Chem.* 43 (3), 340–343, [10.1134/S106816201703013X](https://doi.org/10.1134/S106816201703013X)
  23. Pakhomov AA, Deyev IE, Ratnikova NM, Chumakov SP, Mironiuk VB, Kononevich YN, Muzafarov AM, **Martynov VI** (2017). BODIPY-based dye for no-wash live-cell staining and imaging. *Biotechniques* 63 (2), 77–79, [10.2144/000114577](https://doi.org/10.2144/000114577)
  24. Pakhomov AA, Kononevich YN, Korlyukov AA, **Martynov VI**, Muzafarov AM (2016). Synthesis, crystal structure and optical properties of a new meso-acrylate BODIPY dye. *MENDELEEV COMMUN* 26 (3), 196–198, [10.1016/j.mencom.2016.04.005](https://doi.org/10.1016/j.mencom.2016.04.005)
  25. Pakhomov AA, Kononevich YN, Stukalova MV, Svidchenko EA, Surin NM, Cherkaev GV, Shchegolikhina OI, **Martynov VI**, Muzafarov AM (2016). Synthesis and photophysical properties of a new BODIPY-based siloxane dye. *Tetrahedron Lett* 57 (9), 979–982, [10.1016/j.tetlet.2016.01.059](https://doi.org/10.1016/j.tetlet.2016.01.059)
  26. Pletneva NV, Pletnev S, Pakhomov AA, Chertkova RV, **Martynov VI**, Muslinkina L, Dauter Z, Pletnev VZ (2016). Crystal structure of the fluorescent protein from *Dendronephthya* sp. in both green and photoconverted red forms. *Acta Crystallogr D Struct Biol* 72 (8), 922–932, [10.1107/S205979831601038X](https://doi.org/10.1107/S205979831601038X)
  27. **Martynov VI**, Pakhomov AA, Popova NV, Deyev IE, Petrenko AG (2016). Synthetic Fluorophores for Visualizing Biomolecules in Living Systems. *Acta Naturae* 8 (4), 33–46, [10.32607/20758251-2016-8-4-33-46](https://doi.org/10.32607/20758251-2016-8-4-33-46)
  28. Pakhomov AA, Chertkova RV, **Martynov VI** (2015). PH-sensor properties of a fluorescent protein from *Dendronephthya* sp. *Russ. J. Bioorganic Chem.* 41 (6), 602–606, [10.1134/S1068162015060114](https://doi.org/10.1134/S1068162015060114)
  29. Il'nitskaya EV, Kononevich YN, Muzafarov AM, Rzhavskiy SA, Shadrin IA, Babaev EV, **Martynov VI**, Pakhomov AA (2015). Preparation and application of a BODIPY-labeled probe for a real-time polymerase chain reaction. *Russ. J. Bioorganic Chem.* 41 (4), 451–453, [10.1134/S1068162015040068](https://doi.org/10.1134/S1068162015040068)
  30. Pletneva NV, Pletnev VZ, Souslova E, Chudakov DM, Lukyanov S, **Martynov VI**, Arhipova S, Artemyev I, Wlodawer A, Dauter Z, Pletnev S (2013). Yellow fluorescent protein phiYFPv (*Phialidium*): Structure and structure-based mutagenesis. *Acta Crystallogr D Biol Crystallogr* 69 (6), 1005–1012, [10.1107/S0907444913004034](https://doi.org/10.1107/S0907444913004034)
  31. Pakhomov AA, Tretyakova SA, **Martynov VI** (2012). The influence of chromophore-protein interactions on spectroscopic properties of the yellow fluorescent protein. *Dokl Biochem Biophys* 445 (1), 207–209, [10.1134/S1607672912040072](https://doi.org/10.1134/S1607672912040072)
  32. Pakhomov AA, **Martynov VI** (2011). A method for the determination of the three-dimensional structure of fluorescent proteins based on homology modeling and mass spectrometry. *Russ. J. Bioorganic Chem.* 37 (3), 383–386, [10.1134/S1068162011030137](https://doi.org/10.1134/S1068162011030137)
  33. Pakhomov AA, **Martynov VI** (2011). [3D-structure determination of fluorescent proteins by homology modeling combined with mass spectrometry]. *Bioorg Khim* 37 (3), 429–432.
  34. Pakhomov AA, **Martynov VI** (2011). Probing the structural determinants of yellow fluorescence of a protein from *Phialidium* sp. *Biochem Biophys Res Commun* 407 (1), 230–235, [10.1016/j.bbrc.2011.03.004](https://doi.org/10.1016/j.bbrc.2011.03.004)
  35. Pletneva NV, Pletnev VZ, Lukyanov KA, Gurskaya NG, Goryacheva EA, **Martynov VI**, Wlodawer A, Dauter Z, Pletnev S (2010). Structural evidence for a dehydrated intermediate in green fluorescent protein chromophore biosynthesis. *J Biol Chem* 285 (21), 15978–15984, [10.1074/jbc.M109.092320](https://doi.org/10.1074/jbc.M109.092320)
  36. Pakhomov AA, Tretyakova IA, **Martynov VI** (2010). Posttranslational reactions resulting in a long-wavelength shift in the spectra of asFP595 protein from *Anemonia sulcata*. *Bioorg Khim* 36 (1), 117–121.
  37. Pakhomov AA, Tretyakova YA, **Martynov VI** (2010). Posttranslational reactions that shift spectra of asFP595, a Protein from *Anemonia sulcata*, towards the long-wavelength region. *Russ. J. Bioorganic Chem.* 36 (1), 109–113, [10.1134/S1068162010010127](https://doi.org/10.1134/S1068162010010127)
  38. Pletnev S, Gurskaya NG, Pletneva NV, Lukyanov KA, Chudakov DM, **Martynov VI**, Popov VO, Kovalchuk

- MV, Wlodawer A, Dauter Z, Pletnev V (2009). Structural basis for phototoxicity of the genetically encoded photosensitizer KillerRed. *J Biol Chem* 284 (46), 32028–32039, [10.1074/jbc.M109.054973](https://doi.org/10.1074/jbc.M109.054973)
39. Pakhomov AA, **Martynov VI** (2009). Posttranslational chemistry of proteins of the GFP family. *Biochemistry (Mosc)* 74 (3), 250–259, [10.1134/S000629790903002X](https://doi.org/10.1134/S000629790903002X)
  40. Pakhomov AA, **Martynov VI** (2008). GFP Family: Structural Insights into Spectral Tuning. *Cell Chem Biol* 15 (8), 755–764, [10.1016/j.chembiol.2008.07.009](https://doi.org/10.1016/j.chembiol.2008.07.009)
  41. Pakhomov AA, **Martynov VI** (2007). Chromophore aspartate oxidation-decarboxylation in the green-to-red conversion of a fluorescent protein from *Zoanthus* sp. 2. *Biochemistry* 46 (41), 11528–11535, [10.1021/bi700721x](https://doi.org/10.1021/bi700721x)
  42. Pletneva N, Pletnev V, Tikhonova T, Pakhomov AA, Popov V, **Martynov VI**, Wlodawer A, Dauter Z, Pletnev S (2007). Refined crystal structures of red and green fluorescent proteins from the button polyp *Zoanthus*. *Acta Crystallogr D Biol Crystallogr* 63 (10), 1082–1093, [10.1107/S0907444907042461](https://doi.org/10.1107/S0907444907042461)
  43. Pletneva NV, Pletnev SV, Chudakov DM, Tikhonova TV, Popov VO, **Martynov VI**, Wlodawer A, Dauter Z, Pletnev VZ (2007). Three-dimensional structure of yellow fluorescent protein zYFP538 from *Zoanthus* sp. at the resolution 1.8 angstrom. *Bioorg Khim* 33 (4), 421–430.
  44. Pletneva NV, Pletnev SV, Chudakov DM, Tikhonova TV, Popov VO, **Martynov VI**, Wlodawer A, Dauter Z, Pletnev VZ (2007). Three-dimensional structure of yellow fluorescent protein zYFP538 from *Zoanthus* sp. at the resolution 1.8 Å. *Russ. J. Bioorganic Chem.* 33 (4), 390–398, [10.1134/S1068162007040048](https://doi.org/10.1134/S1068162007040048)
  45. Tretyakova YA, Pakhomov AA, **Martynov VI** (2007). Chromophore structure of the kindling fluorescent protein asFP595 from *Anemonia sulcata*. *J Am Chem Soc* 129 (25), 7748–7749, [10.1021/ja071992c](https://doi.org/10.1021/ja071992c)
  46. Pakhomov AA, Pletneva NV, Balashova TA, **Martynov VI** (2006). Structure and reactivity of the chromophore of a GFP-like chromoprotein from *Condylactis gigantea*. *Biochemistry* 45 (23), 7256–7264, [10.1021/bi060207q](https://doi.org/10.1021/bi060207q)
  47. Pletneva N, Pletnev S, Tikhonova T, Popov V, **Martynov V**, Pletnev V (2006). Structure of a red fluorescent protein from *Zoanthus*, zRFP574, reveals a novel chromophore. *Acta Crystallogr D Biol Crystallogr* 62 (5), 527–532, [10.1107/S0907444906007852](https://doi.org/10.1107/S0907444906007852)
  48. Yampolsky IV, Remington SJ, **Martynov VI**, Potapov VK, Lukyanov S, Lukyanov KA (2005). Synthesis and properties of the chromophore of the asFP595 chromoprotein from *Anemonia sulcata*. *Biochemistry* 44 (15), 5788–5793, [10.1021/bi0476432](https://doi.org/10.1021/bi0476432)
  49. Pakhomov AA, Martynova NY, Gurskaya NG, Balashova TA, **Martynov VI** (2004). Photoconversion of the chromophore of a fluorescent protein from *Dendronephthya* sp. coral. *Biochemistry (Mosc)* 69 (8), 1108–1117.
  50. Pakhomov AA, Martynova NY, Gurskaya NG, Balashova TA, **Martynov VI** (2004). Photoconversion of the chromophore of a fluorescent protein from *Dendronephthya* sp. *Biochemistry (Mosc)* 69 (8), 901–908, [10.1023/B:BIRY.0000040223.09641.29](https://doi.org/10.1023/B:BIRY.0000040223.09641.29)
  51. **Martynov VI**, Maksimov BI, Martynova NY, Pakhomov AA, Gurskaya NG, Lukyanov SA (2003). A Purple-blue Chromoprotein from *Goniopora tenuidens* Belongs to the DsRed Subfamily of GFP-like Proteins. *J Biol Chem* 278 (47), 46288–46292, [10.1074/jbc.M306810200](https://doi.org/10.1074/jbc.M306810200)
  52. Fradkov AF, Verkhusha VV, Staroverov DB, Bulina ME, Yanushevich YG, **Martynov VI**, Lukyanov S, Lukyanov KA (2002). Far-red fluorescent tag for protein labelling. *Biochem J* 368 (1), 17–21, [10.1042/BJ20021191](https://doi.org/10.1042/BJ20021191)
  53. Gurskaya NG, Fradkov AF, Tersikh A, Matz MV, Labas YA, **Martynov VI**, Yanushevich YG, Lukyanov KA, Lukyanov SA (2001). GFP-like chromoproteins as a source of far-red fluorescent proteins. *FEBS Lett* 507 (1), 16–20, [10.1016/S0014-5793\(01\)02930-1](https://doi.org/10.1016/S0014-5793(01)02930-1)
  54. **Martynov VI**, Savitsky AP, Martynova NY, Savitsky PA, Lukyanov KA, Lukyanov SA (2001). Alternative cyclization in GFP-like proteins family. The formation and structure of the chromophore of a purple chromoprotein from *Anemonia sulcata*. *J Biol Chem* 276 (24), 21012–21016, [10.1074/jbc.M100500200](https://doi.org/10.1074/jbc.M100500200)
  55. **Martynov VI**, Kostina MB, Feigina Mlu, Miroshnikov AI (1983). Limited proteolysis studies on molecular organization of bovine rhodopsin in the photoreceptor membrane. 9, 734–745.