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Адрес

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Контакты

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Образование

1996–1998	Пущино	ПущГУ	нейробиолог
1991–1996	Нижний Новгород	ННГУ	биофизик

Работа в ИБХ

Заведующий отделом

2018–наст.вр.	Главный научный сотрудник
2021–наст.вр.	Заместитель директора по науке
2018–2022	Заведующий лабораторией

Членство в советах и комиссиях ИБХ

Ученый совет

Аттестационная комиссия

Членство в сообществах

С 2019 – действительный член Европейской Академии;
С 2017 – член ученого совета Института биоорганической химии;
С 2016 – член-корреспондент Российской Академии Наук;
С 2016 – член диссертационного совета по физиологии/ биофизике (Д 212.166.21)
2014 – 2018 – член ученого совета Нижегородского государственного университета.

Участие в научных обществах

С 2017 – действительный член Физиологического Общества Великобритании;
С 2006 – член Общества Нейронаук, Япония;
С 2003 – член Общества изучения мозга, Финляндия;
С 2002 – 2017 – член Физиологического общества Великобритании;
1999 – 2002 – аффилированный член Физиологического общества Великобритании;
С 1999 – член Общества Нейронаук, США (ID# 100006439).

Степени и звания

2016 Член-корреспондент РАН

Профессор

2002 Доктор наук (Биологические науки)

Гранты и проекты

2023– наст.вр.	Эффект морфологической перестройки астроцитов на функции мозга
2021– 2022	ГФЕН: Влияние диеты с высоким содержанием жиров и сахаров на нейрон-глиальные взаимодействия в мозге
2020– 2022	Влияние сокращения потребляемых калорий на нейрон-глиальные взаимодействия при старении и в модели болезни Альцгеймера
2019– 2019	Международная научная конференция «Baikal Neuroscience Meeting 2019»
2017– 2021	Физиологические изменения в астроцитах при синаптической пластичности и патологических процессах в мозге

Публикации

1. Sergeeva AD, Panova AS, Ivanova AD, Khramova YV, Morozova KI, Kotova DA, Guryleva AV, Khokhlov DD, Kelmanson IV, Vasilev AV, Kostyuk AI, **Semyanov AV**, Oleinikov VA, Belousov VV, Machikhin AS, Brazhe NA, Bilan DS (2024). Where in the tissues of Danio rerio is more H₂O₂ produced during acute hypoxia? *Antioxid Redox Signal* , , [10.1089/ars.2024.0563](https://doi.org/10.1089/ars.2024.0563)
2. Kostyuk AI, Rapota DD, Morozova KI, Fedotova AA, Jappy D, **Semyanov AV**, Belousov VV, Brazhe NA, Bilan DS (2024). Modern optical approaches in redox biology: Genetically encoded sensors and Raman spectroscopy. *J Free Radic Biol Med* 217, 68–115, [10.1016/j.freeradbiomed.2024.03.010](https://doi.org/10.1016/j.freeradbiomed.2024.03.010)
3. Christie IN, Theparambil SM, Braga A, Doronin M, Hosford PS, Brazhe A, Mascarenhas A, Nizari S, Hadjihambi A, Wells JA, Hobbs A, **Semyanov A**, Abramov AY, Angelova PR, Gourine AV (2023). Astrocytes produce nitric oxide via nitrite reduction in mitochondria to regulate cerebral blood flow during brain hypoxia. *Cell Rep* 42 (12), 113514, [10.1016/j.celrep.2023.113514](https://doi.org/10.1016/j.celrep.2023.113514)
4. Popov A, Brazhe N, Morozova K, Yashin K, Bychkov M, Nosova O, Sutyagina O, Brazhe A, Parshina E, Li L, Medyanik I, Korzhevskii DE, Shenkarev Z, Lyukmanova E, Verkratsky A, **Semyanov A** (2023). Mitochondrial malfunction and atrophy of astrocytes in the aged human cerebral cortex. *Nat Commun* 14 (14), 8380, [10.1038/s41467-023-44192-0](https://doi.org/10.1038/s41467-023-44192-0)
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6. Kotova DA, Ivanova AD, Pochechuev MS, Kelmanson IV, Khramova YV, Tiaglik A, Sudoplatov MA, Trifonova AP, Fedotova A, Morozova K, Katrukha VA, Sergeeva AD, Raevskii RI, Pestriakova MP, Solotenkov MA, Stepanov EA, Tsopina AS, Moshchenko AA, Shestopalova M, Zalygin A, Fedotov IV, Fedotov AB, Oleinikov V, Belousov VV, **Semyanov A**, Brazhe N, Zheltikov AM, Bilan DS (2023). Hyperglycemia exacerbates ischemic stroke not through increased generation of hydrogen peroxide. *J Free Radic Biol Med* 208, 153–164, [10.1016/j.freeradbiomed.2023.08.004](https://doi.org/10.1016/j.freeradbiomed.2023.08.004)
7. Barshutina M, Doroshina N, Baizhumanov A, Nikelshparg E, Fedotova A, Popov A, **Semyanov A**, Yakubovsky D, Tselikov G, Luneva O, Kirilyuk I, Maksimov G, Volkov V, Arsenin A, Brazhe N, Novikov S (2023). SERS substrates based on rose petal replicas for the oxidative stress detection. *Appl Surf Sci* 626, , [10.1016/j.apsusc.2023.157281](https://doi.org/10.1016/j.apsusc.2023.157281)
8. Verkratsky A, **Semyanov A** (2023). Decline of astrocyte Ca²⁺ signalling in Alzheimer's disease: STIM1 to the rescue! *Cell Calcium* 113, 102756, [10.1016/j.ceca.2023.102756](https://doi.org/10.1016/j.ceca.2023.102756)
9. Lin SS, Zhou B, Chen BJ, Jiang RT, Li B, Illes P, **Semyanov A**, Tang Y, Verkratsky A (2023). Electroacupuncture prevents astrocyte atrophy to alleviate depression. *Cell Death Dis* 14 (5), 343, [10.1038/s41419-023-05839-4](https://doi.org/10.1038/s41419-023-05839-4)

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11. (книга) Verkhratsky A, **Semyanov A** (2023). Astrocytes in Ageing. *Subcell Biochem* 103, 253–277, [10.1007/978-3-031-26576-1_11](https://doi.org/10.1007/978-3-031-26576-1_11)
12. Xiong XY, **Semyanov A**, Tang Y (2022). Restored oligodendrogenesis by fibroblast growth factor 17: molecular mechanism for rejuvenating ageing-related memory deficit. *Signal Transduct Target Ther* 7 (1), 237, [10.1038/s41392-022-01092-x](https://doi.org/10.1038/s41392-022-01092-x)
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15. Verkhratsky A, **Semyanov A** (2022). The great astroglial metabolic revolution: Mitochondria fuel astrocyte homeostatic support and neuroprotection. *Cell Calcium* 104, 102583, [10.1016/j.ceca.2022.102583](https://doi.org/10.1016/j.ceca.2022.102583)
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33. Verkhratsky A, **Semyanov A**, Zorec R (2020). Physiology of Astroglial Excitability. *Function* 1 (2), zqaa016, [10.1093/function/zqaa016](https://doi.org/10.1093/function/zqaa016)
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38. Popov A, Denisov P, Bychkov M, Brazhe A, Lyukmanova E, Shenkarev Z, Lazareva N, Verkhratsky A, **Semyanov A** (2020). Caloric restriction triggers morphofunctional remodeling of astrocytes and enhances synaptic plasticity in the mouse hippocampus. *Cell Death Dis* 11 (3), 208, [10.1038/s41419-020-2406-3](https://doi.org/10.1038/s41419-020-2406-3)
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44. (конференция), Brazhe AR, **Semyanov AV**, Verkhratsky AN, Denisov P (2019). High-fat (Western) diet induces morphofunctional remodeling of astrocytes in mouse hippocampus. 67 (S1), E399–E400, [10.1002/glia.23675](https://doi.org/10.1002/glia.23675)
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49. (конференция) Kustikova VD, Krivonosov MI, Denisov PA, Zaikin AA, Ivanchenko MV, Meyerov IB, **Semyanov AV** (2018). Time-lapse imaging for calcium activityanalysis in astrocytes with automatic video processing. *Opera Med Physiol* 4, 83.
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